

The Numerical Assessment of Cerebral Blood Flow in Immature Brain of Preterm Infants ^{*} ^{**}

Irina Sidorenko¹[0000-0003-4158-9416], Varvara Turova²[0000-0002-6613-3311],
Andrey Kovtanyuk²[0000-0002-3286-110X], and
Renée Lampe²[0000-0002-1016-0249]

¹ Mathematical Faculty, Chair of Mathematical Modelling,
Technical University of Munich, Garching, Germany
sidorenk@ma.tum.de

² School of Medicine, Klinikum rechts der Isar, Orthopedic Department,
Research Unit for Pediatric Neuroorthopedics and Cerebral Palsy
of the Buhl-Strohmaier Foundation,
Technical University of Munich, Munich, Germany
turova@ma.tum.de, andrei.kovtaniuk@tum.de, renee.lampe@tum.de

Abstract. Intracerebral hemorrhage is the most dangerous complication in the development of premature infants. It is strongly connected with disturbances in cerebral blood flow (*CBF*) and fragility of small blood vessels in germinal matrix (*GM*), which is a highly vascularized layer of the premature brain. Permanent control of *CBF* value and its reaction on changes in mean arterial pressure (*MAP*), arterial carbon dioxide partial pressure (*pCO₂*), and oxygen partial pressure (*pO₂*) are of great importance in the clinical treatment of preterm newborns. The mathematical model for the calculation of *CBF* in immature brain, earlier proposed by Nikolai Botkin and his colleagues, included the dependence of the number of cerebral vessels, their diameter and length on the gestational age, volume of *GM* and brain weight. Furthermore, the vascular response of *CBF* to the change of *MAP* and *pCO₂* was incorporated into the model by increasing or decreasing the diameter of blood vessels (i.e. vasodilation or vasoconstriction). The objective of the present study is modeling of *pO₂* effect on *CBF* in the immature brain of preterm infants by accounting the phenomenological (experimental) dependence of *CBF* on *pO₂* changes. Numerically calculated *CBF* reactivity to changes of *MAP*, *pCO₂* and *pO₂* demonstrates similar values as those observed in experimental studies. The developed mathematical model for *CBF* calculation can be a useful tool in both theoretical research of blood circulation and clinical nursing of preterm infants.

Keywords: Cerebral blood flow · Preterm infant · Immature brain · Germinal matrix · Intracerebral hemorrhage · Mean arterial pressure · Carbon dioxide partial pressure · Oxygen partial pressure.

^{*} Supported by the Klaus Tschira Foundation, Würth Foundation, and Buhl-Strohmaier Foundation

^{**} Copyright ©2020 for this paper by its authors. Use permitted under Creative Commons License Attribution 4.0 International (CC BY 4.0).

1 Introduction

According to the World Health Organization statistics, more than 10% of infants are born preterm, i.e. before 37 completed weeks of gestation, and the preterm birth rate is increasing worldwide [3]. Intracranial hemorrhage (*ICH*) is the major complication of the preterm birth that occurs in 20% to 25% of neonates born before the 30th week of gestation (*WG*) and/or with body weight less than 1500 grams at birth [31]. It often leads to lifelong impairment, such as cerebral palsy, and may cause permanent disorder of the postural and musculoskeletal system, learning disabilities, behavioral problems, speech disorders, perception deficits and seizure disorders. *ICH* typically originates in the germinal matrix (*GM*) [1], which is a highly vascularized area of the developing brain. Volumetric analysis of the germinal matrix provided using 3D MR measurements [20] has shown that the *GM* reaches its maximum size at 23 *WG* and practically disappears by 34 *WG*. A highly fragile microvessel network of *GM* is vulnerable to destruction, which may occur due to the spontaneous fluctuations of cerebral blood flow (*CBF*) caused by the impaired autoregulation [35]. Due to this, the continuous monitoring of *CBF* is important issue in clinical nursing of preterm infants. During the last decades, several techniques, such as near-infrared spectroscopy [12], Xenon-133 clearance measurements [19], transcranial Doppler ultrasonography [27], MRI based arterial spin labeling [11], and diffusion correlation spectroscopy [12] became available for measurements of the entire brain *CBF*. However, none of these techniques are currently used in neonatal clinical practice for the regular monitoring of *CBF*. The numerical assessment of *CBF* using standard clinical records [4, 5, 21, 30] may become a promising approach for clinical applications.

The previous mathematical models [21, 29, 30] evaluated *CBF* from the brain weight (*BW*) estimated from the gestational age (*GA*), mean arterial pressure (*MAP*), and carbon dioxide partial pressure (*pCO₂*) available from the standard clinical records. The arterial oxygen partial pressure (*pO₂*) is another important clinically measured parameter that affects *CBF*. There is evidence [22, 25] that the brain reacts to *pO₂* changes by inverse *CBF* changes, which is described by negative *pO₂* reactivity. Moderate changes in arterial *pO₂* do not influence *CBF* noticeably. Acute hypoxia causes an increase in *CBF* via vasodilation of cerebral arteries and arterioles [14, 26]. This elevation appears to be a threshold phenomenon [22]: *CBF* does not change until *pO₂* falls below 50 mmHg [10, 18], but beneath this limit *CBF* increases substantially [22, 24]. A strong decrease of *pO₂* can increase *CBF* up to 400%, comparing to normoxia condition [7, 18].

While *CBF* is greatly increased by a reduction in *pO₂*, the elevation of *pO₂* level typically causes less pronounced, but regular reduction in *CBF* [34]. The study [26] has shown significant decrease in *CBF* velocity in 15 of the 17 premature infants with hyperoxia during the first few days of life.

The purpose of the present work is accounting for the effect of *pO₂* on *CBF* in the immature brain of preterm infants. The enhancement of the mathematical model for *CBF* calculation [21, 30] is performed by altering the vessels' diameter during hypoxia or hyperoxia as a response to changes in arterial *pO₂* [14, 26].

2 Methods

2.1 Modeling of CBF in the Immature Brain with a Germinal Matrix

A mathematical model for the calculation of *CBF* [21, 30] is based on a hierarchical cerebrovascular model for the adult brain [29] in which the cerebral vascular system is described by 19 levels according to the morphological characteristics of the vessels. Levels from 1 to 9 correspond to arteries and arterioles, level 10 accounts for capillaries, and levels from 11 to 19 simulate venules and veins. To adjust the model to the immature brain, the number of vessels m_j as well as their length l_j and diameter d_j are scaled down on each level j according to the brain weight (BW) of infant as follows:

$$m_j = M_j \cdot (1200/BW - (1200/BW - 1) \cdot |j - 10|/9)^{-1}, \quad (1)$$

$$l_j = L_j \cdot (1 + 0.1 \cdot (1200/BW - 1) \cdot |j - 10|/9)^{-1}, \quad (2)$$

$$d_j = D_j \cdot (1 + 0.1 \cdot (1200/BW - 1) \cdot |j - 10|/9)^{-1}. \quad (3)$$

Here, M_j , L_j and D_j are the number, the length, and the diameter of vessels in level j of the adult brain, respectively, and the value of 1200 g corresponds to the approximate weight of the adult brain [29]. The coefficient 0.1 is used to scale the vessel length and diameter to the experimental measurements [2, 32]. The brain weight of the preterm infant (BW) is computed from the gestational age in weeks (WG) according to the regression formula [16]:

$$BW(WG) = 255.25 - 35.44 \cdot WG + 1.52 \cdot WG^2 - 0.01 \cdot WG^3. \quad (4)$$

Such a model keeps the number of main arteries and veins constant across age and increases the number of arterioles, capillaries and venules according to the amount of the brain tissue, which grows with age. In contrast to the vessel's number, the length and diameter rise with age for large vessels, but remain the same for capillaries.

Anatomic analysis of blood vessels in a germinal matrix [1, 2, 32] have shown, that their morphological and histological characteristics are close to other brain capillaries. Therefore, the presence of *GM* is modeled as an additional parallel circuit on the capillary level ($j = 10$). The number of non-*GM* capillaries in the larger circuit is given by:

$$m_B = M_{10}/(1200/BW) \cdot (1 - GM_{vf}). \quad (5)$$

The number of *GM* capillaries in the smaller circuit is given by:

$$m_{GM} = M_{10}/(1200/BW) \cdot GM_{vf} \cdot 1.5. \quad (6)$$

Here, M_{10} is the number of capillaries on the 10th level of the adult vascular network [29], GM_{vf} is the volume fraction of the germinal matrix relative to the

total brain volume [20], and the factor 1.5 describes a vascular density correction factor [2] for the *GM*. The values of capillary length and diameter are taken from the literature as follows: $l_{GM} = 40 \mu\text{m}$, $d_{GM} = 6.7 \mu\text{m}$ for the *GM* [2, 32], and $l_B = 60 \mu\text{m}$ and $d_B = 5.6 \mu\text{m}$ for the rest of the brain [29].

The total *CBF* is calculated from the Kirchhoff's law as follows:

$$CBF = (MAP - P_{ic}) \cdot \left(\sum_{j=1}^{19} RES_j^{level} \right)^{-1}. \quad (7)$$

Here, P_{ic} is the intracranial pressure taken for preterm infants as 5 mmHg [13] and RES_j^{level} is the total vascular resistance of the level j . All levels except for the capillary one consist of m_j parallel connected vessels with the individual resistance RES_j . Thus, the total resistance can be calculated as follows:

$$RES_j^{level} = RES_j / m_j. \quad (8)$$

The total resistance of the capillary level $j = 10$, consisting of two parallel circuits (*GM* and non-*GM* capillaries), can be calculated as:

$$RES_{10}^{level} = \left((RES_{GM} / m_{GM})^{-1} + (RES_B / m_B)^{-1} \right)^{-1}. \quad (9)$$

The resistances RES_j , RES_B , and RES_{GM} are calculated using a micropolar fluid model [15, 28]. Thus, the conservation of angular momentum results in new equations describing the rotation of fluid particles on the micro-scale [17, 33].

2.2 Accounting for Arterial pO_2

According to the experimental data, the effect of the pO_2 on *CBF* is different for three different states: hypoxia with $pO_2 < 50$ mmHg, normoxia with $50 \text{ mmHg} \leq pO_2 \leq 70$ mmHg, and hyperoxia with $pO_2 > 70$ mmHg. Experimental study [25] has shown that the direct effect of pO_2 on *CBF* in normoxia state was about 10 times less than the effect of pCO_2 , suggesting that the influence of pO_2 on *CBF* in this case is negligible. Therefore, in the mathematical model presented here, the effect of pO_2 in normoxia is neglected. The effect of pO_2 in hyperoxia or hypoxia is modeled by the decrease or increase of the vessels' diameter. An important point is that pO_2 and pCO_2 influence blood circulation independently of each other and have an additive effect on the vessel diameter [14, 26]. In the mathematical model, first the influence of pCO_2 on the reference diameter of the vessel is accounted for, as described in [30], and then the effect of pO_2 is added. The myogenic response to the *MAP* changes is included afterwards as it is specified in [30].

The vasodilation during hypoxia and vasoconstriction during hyperoxia is calculated as a linear increase or decrease of the vessel diameter as:

$$d_{pO_2} = d_{pCO_2} + pv * d_{pCO_2}. \quad (10)$$

Here, d_{pCO_2} is the diameter of the vessel after accounting for the pCO_2 effect as described in [30]. The coefficient pv depends on vessel type and is equal to the relative change of the vessel’s diameter measured in animal experiments [14, 26]. Neither dilations nor constrictions due to pO_2 alterations were detected for capillaries. During hypoxia, a more obvious vasodilation in the veins than in the arteries was observed. In mild hypoxia, with $40 \text{ mmHg} \leq pO_2 < 50 \text{ mmHg}$, an increase from the reference diameter size was 6% for arteries and 9% for veins. Furthermore, in severe hypoxia, with $pO_2 < 40 \text{ mmHg}$, the diameter increased by 20% for arteries and by 34% for veins. While hypoxia caused considerable increases in blood flow, hyperoxia produced only a moderate decrease. During hyperoxia, both arteries and veins constricted slightly and similarly. In mild hyperoxia, with $70 \text{ mmHg} < pO_2 \leq 80 \text{ mmHg}$, a decrease from the reference diameter size was 5% for arteries and 6% for veins, while in severe hyperoxia, with $pO_2 > 80 \text{ mmHg}$, the decrease was 7% both for arteries and for veins.

3 Results and Discussion

The number of vessels on different levels of the hierarchical cerebrovascular model versus gestational age in weeks is shown in Fig. 1. While the number of large arteries ($j = 1$) and veins ($j = 19$) stays constant, the number of smaller vessels ($2 \leq j \leq 18$) increases with gestational age. On capillary level ($j = 10$), the number of non-*GM* capillaries also grows with gestational age, whilst the number of *GM* capillaries rapidly decreases and becomes zero after 33 *WG* (see Fig. 1b). This is in agreement with the observation that the number of *ICH* cases rapidly decreases after 34 *WG* [1].

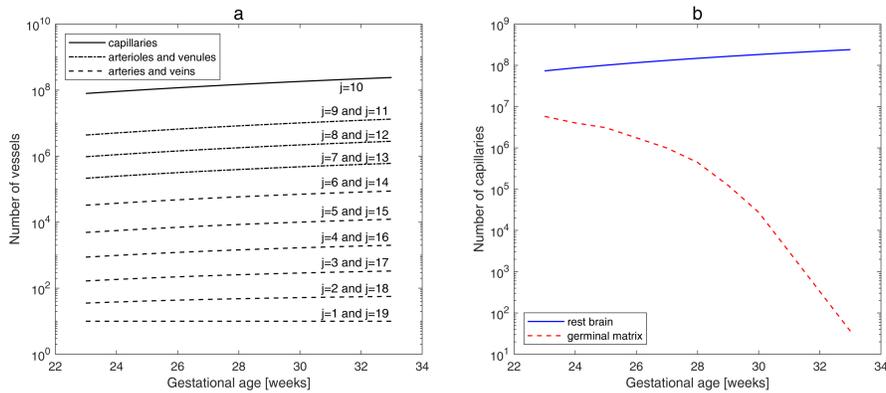


Fig. 1. Number of vessels for different gestational ages on different levels of the hierarchical cerebral vascular model (a) and on the capillary level (b).

The initial vessel diameter slowly increases with gestational age (see Fig. 2a). The CBF reactivity on changes in pO_2 , pCO_2 , and MAP is regulated by the vascular activity, i.e. vasoconstriction and vasodilation, which is modeled as the alteration of the vessel's diameter. The effect of pO_2 changes on diameters of the largest arteries and veins is demonstrated in Fig. 2b. The most considerable alteration of the vessel's diameter is observed for veins during hypoxia. The diameter of the largest veins increases from 1.53 mm to 2.1 mm at age 23 WG and from 2.37 mm to 3.2 mm at age 33 WG .

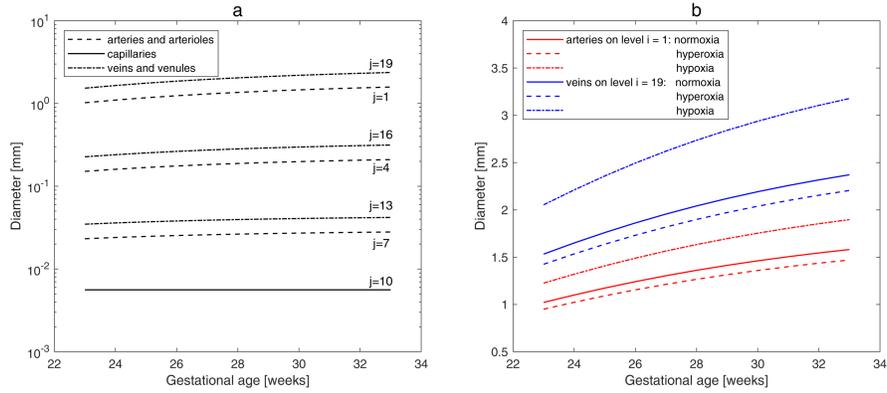


Fig. 2. Initial diameter of vessels for different gestational ages on different levels of the hierarchical cerebral vascular model (a) and at different values of pO_2 (b).

The dependence of CBF on changes in pO_2 , pCO_2 , and MAP is demonstrated in Fig. 3. The CBF stays constant for pO_2 values between 50 mmHg and 70 mmHg, as it has been described in [10, 18]. In hypoxia condition, CBF demonstrates a considerable increase with a threshold phenomenon at $pO_2 = 50$ mmHg described in [18, 22]. At $pO_2 = 50$ mmHg, CBF starts to increase with decreasing pO_2 , reaching a twofold elevation of CBF value with respect to normoxia level (see Fig. 3a) at $pO_2 = 30$ mmHg, as it has been observed in [8, 18]. In hyperoxia condition, the increase of pO_2 from 70 mmHg to 80 mmHg causes the 20% decrease in CBF (see Fig. 3a). Such a reactivity is in good agreement with experimental studies [6, 8, 23], where CBF reactivity of 10–30% per 1 kPa (7.5 mmHg) increase of arterial pO_2 was measured in preterm infants.

Simultaneous dependence of CBF on MAP , pCO_2 , and pO_2 is shown on Figure 3b. Whilst the CBF reactivity on pCO_2 changes has linear behavior, the CBF reactivity on MAP demonstrates a plateau which corresponds to the cerebral autoregulation observed in experiments [1, 9]. The calculated values of CBF and its reactivity to changes in main medical parameters are in good agreement with experimental measurements presented in the literature [1, 6, 8–10, 18, 22, 23]. Thus, the mathematical model developed provides a realistic description

of physiological processes and can be proposed as a useful tool both for the theoretical research of cerebral circulation and the clinical nursing of preterm infants.

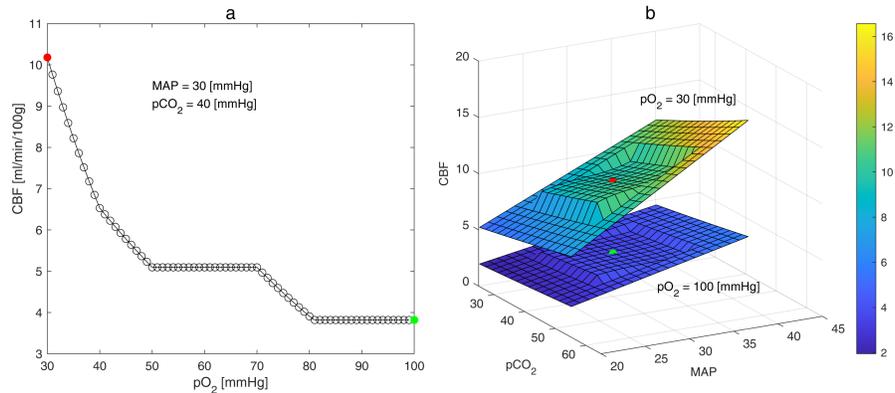


Fig. 3. Dependence of *CBF* on pO_2 (a), pCO_2 and *MAP* (b). Both plots correspond to the 23 *WG*. The red and green points in both plots correspond to the same values of pO_2 , pCO_2 and *MAP*.

References

1. Ballabh, P.: Intraventricular hemorrhage in premature infants: mechanism of disease. *Pediatr. Res.* **67**(1), 1–8 (2010). <https://doi.org/10.1203/PDR.0b013e3181c1b176>
2. Ballabh, P., Braun, A., Nedergaard, M.: Anatomic analysis of blood vessels in germinal matrix, cerebral cortex, and white matter in developing infants. *Pediatr. Res.* **56**(1), 117–24 (2004). <https://doi.org/10.1203/01.PDR.0000130472.30874.FF>
3. Blencowe, H., Cousens, S., Chou, D., Oestergaard, M., Say, L., Moller, A.B., Kinney, M., Lawn, J.: Born too soon: the global epidemiology of 15 million preterm births. *Reprod. Health.* **10**, S2 (2013). <https://doi.org/10.1186/1742-4755-10-S1-S2>
4. Botkin, N., Turova, V., Lampe, R.: Feedback control of impaired cerebral autoregulation in preterm infant: Mathematical modelling. In: 25th Mediterranean Conference on Control and Automation (MED), pp. 229–234. IEEE, Valletta (2017). <https://doi.org/10.1109/MED.2017.7984123>
5. Botkin, N.D., Turova, V.L., Kovtanyuk, A.E., Sidorenko, I.N., Lampe R.: Extended model of impaired cerebral autoregulation in preterm infants: Heuristic feedback control. *Math. Biosci. Eng.* **16**(4), 2334–2352, (2019). <https://doi.org/10.3934/mbe.2019117>
6. Brew, N., Walker, D., Wong, F. Y.: Cerebral vascular regulation and brain injury in preterm infants. *Am. J. Physiol. Reg., Integr. Compar. Physiol.* **306**(11), R773–R786 (2014). <https://doi.org/10.1152/ajpregu.00487.2013>

7. Cipolla, M.J.: The cerebral circulation. *Colloquium Series on Integrated Systems Physiology: From Molecule to Function*, **1**(1), 1–59 (2009). <https://doi.org/10.4199/C00005ED1V01Y200912ISP002>
8. Cold, G.E., Dahl, B.L.: *Topics in neuroanaesthesia and neurointensive care: Experimental and clinical studies upon cerebral circulation, metabolism and intracranial pressure*. Springer, Berlin, Heidelberg (2002). <https://doi.org/10.1007/978-3-662-04845-0>
9. Da Costa, C.S., Czosnyka, M., Smielewski, P., Mitra, S., Stevenson, G.N., Austin, T.: Monitoring of cerebrovascular reactivity for determination of optimal bloodpressure in preterm infants. *J. Pediatr.* **167**(1), 86–91 (2015). <https://doi.org/10.1016/j.jpeds.2015.03.041>
10. Deorari, A.K.: *Blood gas analysis*. Deorari, AIIMS (2008)
11. De Vis, J.B., Hendrikse, J., Groenendaal, F., De Vries, L.S., Kersbergen, K.J., Benders, M.J., Petersen, E.T.: Impact of neonate haematocrit variability on the longitudinal relaxation time of blood: Implications for arterial spin labelling MRI. *Neuroimage. Clin.* **4**, 517–525 (2014). <https://doi.org/10.1016/j.nicl.2014.03.006>
12. Diop, M., Kishimoto, J., Toronov, V., Lee, D.S., Lawrence, K.S.: Development of a combined broadband near-infrared and diffusion correlation system for monitoring cerebral blood flow and oxidative metabolism in preterm infants. *Biomed. Opt. Express.* **6**(10), 3907–18 (2015). <https://doi.org/10.1364/BOE.6.003907>
13. Easa, D., Tran, A., Bingham, W.: Noninvasive intracranial pressure measurement in the newborn: an alternate method. *Am. J. Dis. Child.* **137**(4), 332–335 (1983). <https://doi.org/10.1001/archpedi.1983.02140300014004>
14. Eperon, G., Johnson, M., David, N.J.: The effect of arterial PO₂ on relative retinal blood flow in monkeys. *Investi. Ophthalmol.* **14**(5), 342–352 (1975)
15. Erdogan, M.E.: Polar effects in the apparent viscosity of a suspension. *Rheol. Acta.* **9**(3), 434–438 (1970). <https://doi.org/10.1007/BF01975413>
16. Guihard-Costa, A.M., Larroche, J.C.: Differential growth between the fetal brain and its infratentorial part. *Early Hum. Dev.* **23**(1), 27–40 (1990). [https://doi.org/10.1016/0378-3782\(90\)90126-4](https://doi.org/10.1016/0378-3782(90)90126-4)
17. Hoffmann, K.-H., Marx, D., Botkin, N.D.: Drag on spheres in micropolar fluids with non-zero boundary conditions for microrotations. *J. Fluid Mech.* **590**, 319–330 (2007). <https://doi.org/10.1017/S0022112007008099>
18. Johnston, A.J., Steiner, L.A., Gupta, A.K., Menon D.K.: Cerebral oxygen vasoreactivity and cerebral tissue oxygen reactivity. *Br. J. Anaesth.* **90**(6), 774–786 (2003). <https://doi.org/10.1093/bja/aeg104>
19. Jayasinha, D., Gill, A.B., Levene, M.I.: CBF reactivity in hypotensive and normotensive preterm infants. *Pediatr Res.* **54**(6), 848–853 (2003). <https://doi.org/10.1203/01.PDR.0000088071.30873.DA>
20. Kinoshita, Y., Okudera, T., Tsuru, E., Yokota, A.: Volumetric analysis of the germinal matrix and lateral ventricles performed using MR images of postmortem fetuses. *Am. J. Neuroradiol.* **22**(2), 382–388 (2001)
21. Lampe, R., Botkin, N., Turova, V., Blumenstein, T., Alves-Pinto, A.: Mathematical modelling of cerebral blood circulation and cerebral autoregulation: towards preventing intracranial hemorrhages in preterm newborns. *Comput. Math. Methods. Med.* **2014**, 965275 (2014). <https://doi.org/10.1155/2014/965275>
22. Lassen, N., A.: Control of cerebral circulation in health and disease. *Circ. Res.* **34**(6), 749–760 (1974). <https://doi.org/10.1161/01.RES.34.6.749>
23. Leahy, F. A., Cates, D., MacCallum, M., Rigatto, H.: Effect of CO₂ and 100% O₂ on cerebral blood flow in preterm infants. *J. Appl. Physiol.* **48**(3), 468–472 (1980). <https://doi.org/10.1152/jappl.1980.48.3.468>

24. Masamoto, K., Tanishita, K.: Oxygen transport in brain tissue. *J. Biomech. Eng.* **131**(7), 074002 (2009). <https://doi.org/10.1115/1.3184694>
25. Menke, J., Michel, E., Rabe, H., Bresser, B. W., Grohs, B., Schmitt, R. M., Jorch, G.: Simultaneous influence of blood pressure, PCO₂, and PO₂ on cerebral blood flow velocity in preterm infants of less than 33 weeks' gestation. *Pediatr. Res.* **34**(2), 173–177 (1993). <https://doi.org/10.1203/00006450-199308000-00014>
26. Nijima, S., Shortland, D.B., Levene, M.L., Evans, D.H.: Transient hyperoxia and cerebral blood flow velocity in infants born prematurely and at full term. *Arch. Dis. Child.* **63**(10 Spec No), 1126–1130 (1988). https://doi.org/10.1136/adc.63.10_spec_no.1126
27. Noori, S., Anderson, M., Soleymani, S., Seri, I.: Effect of carbon dioxide on cerebral blood flow velocity in preterm infants during postnatal transition. *Acta Paediatr.* **103**(8), e334–e339 (2014). <https://doi.org/710.1111/apa.12646>
28. Papautsky, I., Brazzle, J., Ameel, T., Frazierac, A.B.: Laminar fluid behavior in microchannels using micropolar fluid theory. *Sens. Actuat. A. Phys.* **73**(1-2), 101–108 (1999). [https://doi.org/710.1016/S0924-4247\(98\)00261-10](https://doi.org/710.1016/S0924-4247(98)00261-10)
29. Piechnik, S.K., Chiarelli, P.A., Jezzard, P.: Modelling vascular reactivity to investigate the basis of the relationship between cerebral blood volume and flow under CO₂ manipulation. *Neuroimage* **39**(1), 107–118 (2008). <https://doi.org/10.1016/j.neuroimage.2007.08.022>
30. Sidorenko, I., Turova, V., Botkin, N., Eckardt, L., Alves-Pinto, A., Felderhoff-Müser, U., Rieger-Fackeldey, E., Kovtanyuk, A., Lampe, R.: Modeling cerebral blood flow dependence on carbon dioxide and mean arterial blood pressure in the immature brain with accounting for the germinal matrix. *Front. Neurol.* **9**, 812 (2018). <https://doi.org/10.3389/fneur.2018.00812>
31. Schmid, M.B., Reister, F., Mayer, B., Hopfner, R.J., Fuchs, H., Hummler, H.D.: Schmid, M.B., Reister, F., Mayer, B., Hopfner, R.J., Fuchs, H., Hummler, H.D.: Prospective risk factor monitoring reduces intracranial hemorrhage rates in preterm infants. *Dtsch. Arztebl. Int.* **110**(29-30), 489–496 (2013). <https://doi.org/10.3238/arztebl.2013.0489>
Prospective risk factor monitoring reduces intracranial hemorrhage rates in preterm infants. *Dtsch. Arztebl. Int.* **110**(29-30), 489–496 (2013). <https://doi.org/10.3238/arztebl.2013.0489>
32. Trommer, B.L., Groothuis, D.R., Pasternak, J.F.: Quantitative analysis of cerebral vessels in the newborn puppy: the structure of germinal matrix vessels may predispose to hemorrhage. *Pediatr. Res.* **22**(1), 23–28 (1987). <https://doi.org/10.1203/00006450-198707000-00007>
33. Turova, V., Botkin, N., Alves-Pinto, A., Blumenstein, T., Rieger-Fackeldey, E., Lampe, R.: Modelling autoregulation of cerebral blood flow using viability approach. In: *International Symposium on Dynamic Games and Applications*. **15**, 345–363. Birkhäuser, Cham (2016). https://doi.org/10.1007/978-3-319-70619-1_16
34. Ursino, M., Di Giammarco, P., Belardinelli, E.: A mathematical model of cerebral blood flow chemical regulation. I. Diffusion processes. *IEEE Trans. Biomed. Eng.* **36**(2), 183–191 (1989). <https://doi.org/10.1109/10.16465>
35. Wong, F.Y., Leung, T.S., Austin, T., Wilkinson, M., Meek, J.H., Wyatt, J.S., Walker, A.M.: Impaired autoregulation in preterm infants identified by using spatially resolved spectroscopy. *Pediatrics* **121**(3), e604–e611 (2008). <https://doi.org/10.1542/peds.2007-1487>